



## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

<b>(51) International Patent Classification 7 :</b> <b>C12Q 1/68, G01N 30/72, 33/50</b>	<b>A2</b>	<b>(11) International Publication Number:</b> <b>WO 00/31300</b> <b>(43) International Publication Date:</b> 2 June 2000 (02.06.00)
<b>(21) International Application Number:</b> PCT/US99/27523 <b>(22) International Filing Date:</b> 22 November 1999 (22.11.99) <b>(30) Priority Data:</b> 09/198,340 24 November 1998 (24.11.98) US <b>(71) Applicant (for all designated States except US):</b> THE JOHNS HOPKINS UNIVERSITY [US/US]; Suite 906, 111 Market Place, Baltimore, MD 21202 (US). <b>(72) Inventors; and</b> <b>(75) Inventors/Applicants (for US only):</b> LAKEN, Steven, J. [US/US]; 38 Lawrence Street, Pepperell, MA 01463 (US). VOGELSTEIN, Bert [US/US]; 3700 Breton Way, Baltimore, MD 21208 (US). KINZLER, Kenneth, W. [US/US]; 1403 Halkirk Way, BelAir, MD 21015 (US). GROOPMAN, John, D. [US/US]; 3744 Birchmere Court, Owings Mills, MD 21117 (US). JACKSON, Peta, E. [GB/US]; 721 S. Ann Street, Baltimore, MD 21231 (US). FRIESEN, Marlin, D. [US/US]; 22, rue Benoit Berlioz, Chassieu (FR). <b>(74) Agents:</b> KAGAN, Sarah, A. et al.; Banner & Witcoff, Ltd., 11th floor, 1001 G Street, N.W., Washington, DC 20001-4597 (US).		<b>(81) Designated States:</b> AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).  <b>Published</b> <i>Without international search report and to be republished upon receipt of that report.</i>
<b>(54) Title:</b> GENOTYPING BY MASS SPECTROMETRIC ANALYSIS OF SHORT DNA FRAGMENTS  <b>(57) Abstract</b>  Genotyping can be accomplished by analysis of short, defined DNA segments using electrospray ionization mass spectrometry. The DNA segments are produced using specially designed primers to amplify a cDNA or genomic DNA template. The primers contain a recognition site for a restriction endonuclease. The amplification products are digested with the restriction endonuclease. Single nucleotide polymorphisms can be detected rapidly and reliably.		

**FOR THE PURPOSES OF INFORMATION ONLY**

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AL	Albania	ES	Spain	LS	Lesotho	SI	Slovenia
AM	Armenia	FI	Finland	LT	Lithuania	SK	Slovakia
AT	Austria	FR	France	LU	Luxembourg	SN	Senegal
AU	Australia	GA	Gabon	LV	Latvia	SZ	Swaziland
AZ	Azerbaijan	GB	United Kingdom	MC	Monaco	TD	Chad
BA	Bosnia and Herzegovina	GE	Georgia	MD	Republic of Moldova	TG	Togo
BB	Barbados	GH	Ghana	MG	Madagascar	TJ	Tajikistan
BE	Belgium	GN	Guinea	MK	The former Yugoslav Republic of Macedonia	TM	Turkmenistan
BF	Burkina Faso	GR	Greece	ML	Mali	TR	Turkey
BG	Bulgaria	HU	Hungary	MN	Mongolia	TT	Trinidad and Tobago
BJ	Benin	IE	Ireland	MR	Mauritania	UA	Ukraine
BR	Brazil	IL	Israel	MW	Malawi	UG	Uganda
BY	Belarus	IS	Iceland	MX	Mexico	US	United States of America
CA	Canada	IT	Italy	NE	Niger	UZ	Uzbekistan
CF	Central African Republic	JP	Japan	NL	Netherlands	VN	Viet Nam
CG	Congo	KE	Kenya	NO	Norway	YU	Yugoslavia
CH	Switzerland	KG	Kyrgyzstan	NZ	New Zealand	ZW	Zimbabwe
CI	Côte d'Ivoire	KP	Democratic People's Republic of Korea	PL	Poland		
CM	Cameroon	KR	Republic of Korea	PT	Portugal		
CN	China	KZ	Kazakhstan	RO	Romania		
CU	Cuba	LC	Saint Lucia	RU	Russian Federation		
CZ	Czech Republic	LI	Liechtenstein	SD	Sudan		
DE	Germany	LK	Sri Lanka	SE	Sweden		
DK	Denmark	LR	Liberia	SG	Singapore		
EE	Estonia						

## **GENOTYPING BY MASS SPECTROMETRIC ANALYSIS OF SHORT DNA FRAGMENTS**

The U.S. Government retains certain rights in this invention due to funding of grant CA43460 awarded by the National Institutes of Health.

### **5      TECHNICAL FIELD OF THE INVENTION**

The invention is related to the area of genome analysis. In particular it is related to the field of detection of genetic polymorphisms.

### **BACKGROUND OF THE INVENTION**

10      One of the most important results of the revolution in genomics research has been the elucidation of genetic variants associated with specific human diseases. Recent examples include variants in BRCA genes predisposing to breast cancer, a variant in Apo E predisposing to dementia, and a variant in prothrombin predisposing to bleeding disorder (1-3). All of these variations are found at relatively high frequencies in certain populations, and testing for the presence of  
15      such mutations can provide critical diagnostic information for management of patients and their families.

20      The discoveries of such variations have stimulated efforts to design approaches for assessing their presence in DNA from clinical samples. Three factors are particularly important for the success of such efforts: accuracy, throughput, and cost. For the evaluation of an individual (or a few) variants, throughput and cost are not generally limiting, but accuracy remains a continuing

concern. Procedures that work well in a research environment are not necessarily appropriate for clinical application, as even a minute fraction of errors in the latter setting can have catastrophic consequences.

5 Many of the methods currently used for variant analysis employ hybridization with specific oligonucleotide probes that can discriminate between the wild-type and variant sequences. Such hybridizations can occur on filters, chips, gels, or in solution (4). Though generally reliable and useful, hybridization techniques suffer from their qualitative, rather than quantitative, nature; most probes will hybridize to all sequence variants at temperatures slightly below the discrimination optimum. The fact that the extent of hybridization of allele-specific oligonucleotides (ASO) is dependent both on the nature of the variation and the surrounding sequences can make ASO difficult to apply without substantial optimization.

15 Among the other strategies for genotyping variants, those that employ mass spectrometry (MS) have received particular attention. MS represents an improvement over gel-based and hybridization systems because the mass spectrometer yields precise information on the molecular mass of the analyte, the procedure can be fully automated, and both DNA strands can be analyzed in parallel. MS can directly assess the nature of polymerase chain reaction (PCR) products themselves, while other techniques only indirectly assess such PCR products, either through hybridization probes (as in ASO) or DNA polymerase-generated methods that use PCR products as templates (4). Such indirect methods introduce additional sources of error into the assays.

25 The feasibility of MS analysis of polymorphic PCR products has been demonstrated (5). However, one limiting factor for analysis of single nucleotide polymorphisms (SNPs) is the mass resolution required for measuring a small difference (9 Da between A and T) in PCR-generated fragments, which are generally on the order of 100 bp long. Procedures have been developed which use PCR products as templates to which peptide nucleic acid probes are hybridized and can then be analyzed by MS (6). This clever technique appears to be useful,

30

but it fails to employ one of the strengths of MS in that the analysis of PCR products is not direct.

There is a continuing need in the art for methods which employ MS but employ direct analysis of amplification products.

5      **SUMMARY OF THE INVENTION**

It is an object of the invention to provide reagents and methods for analyzing genotypes by mass spectrometry. These and other objects of the invention are provided by one or more of the embodiments described below.

10      One embodiment of the invention provides an isolated primer for amplifying a segment of DNA. The primer comprises a linear oligonucleotide consisting of at least 35 nucleotides. The oligonucleotide comprises a 5' end and a 3' end. A first portion of the oligonucleotide consists of at least 13 nucleotides at the 5' end of the oligonucleotide. A second portion of the oligonucleotide consists of from 5 to 22 nucleotides at the 3' end of the oligonucleotide. The first and second portions  
15      of the oligonucleotide are either precisely complementary or substantially complementary to a first portion and a second portion, respectively, of a segment of a cDNA or genomic DNA. Four to eight nucleotides between the first portion and the second portion of the oligonucleotide comprise a recognition site for a restriction endonuclease that cleaves at least five nucleotides removed from its  
20      recognition site. The segment of the cDNA or genomic DNA does not comprise the recognition site.

Another embodiment of the invention provides a pair of purified primers for amplifying a segment of cDNA or genomic DNA. Each primer comprises a linear oligonucleotide consisting of at least 35 nucleotides. A first portion of the  
25      oligonucleotide of at least 13 nucleotides at the 5' end and a second portion of the oligonucleotide of from 5 to 22 nucleotides at the 3' end are either precisely complementary or substantially complementary to a first portion and a second portion of a cDNA or genomic DNA. Between the first portion and the second portion of the oligonucleotide are 4-8 nucleotides which comprise a recognition  
30      site for a restriction endonuclease that cleaves at least five nucleotides from its

recognition site. The segment of the cDNA or genomic DNA does not comprise the recognition site for the restriction endonuclease. Each primer of the pair of primers is complementary to an opposite strand of a double stranded cDNA or genomic DNA molecule. The pair of primers is complementary to two  
5 non-contiguous portions of the double stranded cDNA or genomic DNA molecule, such that 1 to 20 nucleotides separate the two non-contiguous portions of the double stranded cDNA or genomic DNA molecule.

Still another embodiment of the invention provides a kit comprising a plurality of pairs of primers as described in the preceding paragraph.

10 Yet another embodiment of the invention provides a method for producing a short segment of DNA, suitable for analysis by MS. The method comprises the steps of amplifying cDNA or genomic DNA using the pair of primers described above to form amplified DNA and digesting the amplified DNA with the restriction endonuclease to form a short segment of DNA.

15 A further embodiment of the invention provides a method for analyzing a first short segment of DNA comprising a first polymorphic nucleotide to distinguish the first short segment of DNA from a second short segment of DNA comprising a second polymorphic nucleotide. The method comprises the step of applying a mixture of DNA segments to an electrospray ionization/ mass  
20 spectrometer, whereby the DNA segments are denatured and the denatured segments are separated. The mixture of DNA segments is made by the process of amplifying cDNA or genomic DNA of a subject using the pair of primers described above to form amplified DNA and digesting the amplified DNA with the restriction endonuclease to form a short segment of DNA.

25 The invention thus provides the art with novel tools and methods for analyzing the genotype of living organisms, including humans, by electrospray ionization mass spectrometry.

**BRIEF DESCRIPTION OF THE DRAWINGS**

Figure 1 displays the general strategy for the preparation of DNA suitable for short oligonucleotide mass analysis (SOMA). A template is PCR-amplified with primers containing an artificial *BpmI* restriction endonuclease sequence (CTGGAG) embedded within sequences perfectly complementary to the genomic region of interest. The PCR product is digested with *BpmI*, and the internal (interrogated) sequence released by *BpmI* digestion is analysed by the mass spectrometer.

Figures 2A, 2B, and 2C illustrate full-scan electrospray mass spectra of 15-mer oligonucleotide standards corresponding to the antisense strands of the APC codon 1307 AAA allele (Figure 2A) and ATA allele (Figure 2B). The mass that is the most amenable to detection by the mass spectrometer is the  $[M-3H]^3$  peak corresponding to a  $m/z$  of 1519.3 and 1522.3 for the AAA and ATA alleles, respectively. Figure 2C shows the electrospray mass spectrum for the simultaneous ESI-MS analysis of these two oligonucleotide standards, showing baseline separation for the two  $[M-3H]^3$  ions.

Figures 3A and 3B demonstrate ESI-MS analysis of APC codon 1307 variants. The four mass chromatograms for each patient represent the AAA sense (s) mass, the AAA antisense (as) mass, the ATA sense (s) mass and the ATA antisense (as) mass, respectively. The patient in Figure 3A has the ATA/ATA homozygous genotype, while that in Figure 3B has the ATA/AAA heterozygous genotype.

Figures 4A, 4B, and 4C demonstrate ESI-MS/MS SRM analysis of APC codon 1493 variants. Mass chromatograms obtained from genomic DNA of patients with the ACG/ACA, ACA/ACA, and ACG/ACG genotypes, respectively, are presented in Figure 4A, Figure 4B, and Figure 4C, respectively. The masses representing the sense (s) and antisense (as) *BpmI* fragments corresponding to the variant sequences are indicated.

Figures 5A and 5B demonstrate simultaneous analysis of three different APC variants for two patients. For each patient, PCR products containing APC codons 486, 545, and 1756 were combined and introduced into the mass

spectrometer via the HPLC. The sense (s) and antisense (as) signals are indicated for each genotype. Figure 5A represents an individual homozygous at each of the analyzed codons, and Figure 5B was from an individual homozygous for the other allele at each of these codons.

5      **DETAILED DESCRIPTION OF THE INVENTION**

          The present invention provides a method of genotype analysis in which short, defined fragments of amplification products are produced by simple enzymatic digestion and directly analyzed by electro-spray ionization mass spectrometry (ESI-MS). The method, called SOMA (Short Oligonucleotide Mass  
10      Analysis), is simple to implement, extremely accurate, and applicable to most DNA variations.

          The SOMA technique utilizes short DNA segments of defined length. The segments are produced by amplification of a segment of cDNA or genomic DNA of approximately 50, 60, 70, 80, 90, 100, 110, 120, 130, 140, or 150 bp,  
15      preferably about 100 bp, using specially designed amplification primers. The cDNA or genomic DNA can be isolated from a subject organism by methods known in the art. The subject organism can be any organism, for example a human or other animal, a plant, a fungus, or a microorganism such as a bacterium or a virus.

20      Primers can be either precisely complementary or substantially complementary to two non-contiguous portions of the segment of cDNA or genomic DNA. The term "precisely complementary" as used herein refers to nucleic acids that are complementary at every base pair. Thus, a primer is precisely complementary to its template sequence if every nucleotide of the primer  
25      is complementary to every corresponding nucleotide of the template sequence. The term "substantially complementary" refers to nucleotide sequences which are at least 90% identical to their corresponding template sequences as determined by the Smith-Waterman homology search algorithm as implemented in the MPSRCH program (Oxford Molecular) using an affine gap search with a gap open penalty of  
30      12 and a gap extension penalty of 1.



The two non-contiguous portions of the cDNA or genomic DNA to which the primers are complementary flank the portion of the cDNA or genomic DNA containing the polymorphism. The two non-contiguous portions are separated from each other by 1 to about 10, 12, 14, 16, 18, 20, 22, 24, 26, 28, 30, 35, or 40 bp, and preferably by 1 to 20 bp. The two primers are complementary to opposite strands of the cDNA or genomic DNA, such that amplification produces a segment of cDNA or genomic DNA which contains the polymorphism to be analyzed flanked by the primer sequences.

The primer can be a linear oligonucleotide comprising at least 20, 25, 30, 35, 40, 45, 50, 60, or 70 nucleotides, preferably comprising at least 35 nucleotides, and more preferably consisting of from 41 to 44 nucleotides. The primer can comprise a first portion at its 5' end, which comprises at least 8, 9, 10, 11, 12, 13, 14, 15, 16, 18, 20, 22, or 25 nucleotides. Preferably the first portion comprises at least 13 nucleotides. More preferably the first portion consists of from 21 to 22 nucleotides. The first portion of the primer is complementary, or substantially so, to one strand of the cDNA or genomic DNA segment. The primer can also comprise a second portion at its 3' end, which consists of at least 3, 4, 5, 6, 7, 8, or 10 to 18, 19, 20, 21, 22, 23, 24, 26, 28, or 30 nucleotides. Preferably the second portion consists of from 5 to 22 nucleotides, and more preferably the second portion consists of from 14 to 16 nucleotides. The second portion of the primer is complementary, or substantially so, to a second portion of the same strand of the cDNA or genomic DNA segment to which the first primer portion is complementary.

The first and second portions of the primer are separated by a sequence consisting of from 3, 4, or 5 to 7, 8, 9, or 10 nucleotides. Preferably the separating sequence consists of from 4 to 8 nucleotides. The separating sequence comprises a restriction endonuclease recognition sequence. A "restriction endonuclease" or "restriction enzyme" is a bacterial enzyme that binds to a specific recognition site on a double stranded DNA molecule and cleaves the molecule at a specific cleavage site. The "recognition site" is a nucleotide sequence within the double stranded DNA molecule to which the endonuclease

binds. The "cleavage site" is the position at which the endonuclease cuts the double stranded DNA molecule. The position of the cleavage site is relative to the recognition site and is a characteristic of the endonuclease.

5 The restriction endonuclease whose recognition sequence is used is a restriction endonuclease that cleaves at a site distinct from the recognition sequence. The restriction endonuclease can be, for example, a Type IIS restriction endonuclease such as BpmI, BsgI, BseRI, or BciVI. Type IIS restriction endonucleases have asymmetric recognition sites and cleave at a specific distance of up to 20 bp outside their recognition site (20). Using a restriction endonuclease  
10 that cleaves outside the primer is advantageous, because the product of endonuclease treatment can then be a smaller DNA segment than if the endonuclease cleaved within the primer, and a smaller DNA segment enhances the sensitivity of the method. The restriction endonuclease should have a cleavage site distal from its recognition site by at least 3, 4, 5, 6, 8, 10, 12, or 15  
15 nucleotides, and preferably by at least 8 nucleotides. Preferably, the restriction endonuclease recognition sequence will not be found within the amplified segment of cDNA or genomic DNA.

The two portions of the cDNA or genomic DNA which are complementary to the first and second portions of the primer can be separated by from 0, 1, 2, 3,  
20 or 4 nucleotides to 8, 9, 10, 11, 12, 13, 14, 15, or 16 nucleotides. Preferably they are separated by from 4 to 8 nucleotides and more preferably they are separated by 6 nucleotides.

A pair of such primers as described above which flank a segment of cDNA or genomic DNA containing a polymorphism can be used to amplify the  
25 polymorphism. Each primer of the pair is complementary to a different strand of the cDNA or genomic DNA. Therefore, if a first primer of a pair is complementary to the coding strand of the cDNA or genomic DNA segment, then the other primer of the pair must be complementary to the non-coding strand, *i.e.*, the opposite strand, of the cDNA or genomic DNA segment to be amplified. In  
30 this way, when amplification is performed using the pair of primers, the resulting

amplified DNA will contain a copy of the segment of cDNA or genomic DNA between the portions complementary to the primers (Fig. 1).

5 The region of cDNA or genomic DNA containing the polymorphism between the primer-complementary portions can vary in length from 1, 2, 3, 4, or 5 bp to about 16, 18, 20, 22, 24, 26, 30, 35, or 40 bp, but preferably is in the range from 1 to 20 bp. The length of this region is determined by several factors relating to the design of the primer pair used for amplification. Those factors include the composition and length of the portions of cDNA or genomic DNA to which the primers are complementary and the distance between the recognition  
10 and cleavage sites of the restriction endonuclease. Generally, use of shorter segments of cDNA or genomic DNA yield greater mass resolution and greater sensitivity.

Primers according to the invention can be synthesized by any method known in the art for oligonucleotide synthesis. For example, solid phase oligonucleotide  
15 synthesis can be performed by sequentially linking 5' blocked nucleotides to a nascent oligonucleotide attached to a resin, followed by oxidizing and unblocking to form phosphate diester linkages (21). Primers according to the invention are isolated. The term "isolated" as used herein refers to a molecule that is substantially free of undesired contaminants, such as molecules having other  
20 sequences.

Primers of the invention can be made available as a kit. A kit contains, in one or more divided or undivided vessels, a plurality of primers for use in analyzing one or more specific polymorphisms. The primers in a kit are designed to be used together, for example, in pairs which are complementary to regions of a  
25 cDNA or genomic DNA which flank a particular polymorphism. A kit can optionally contain the restriction endonuclease whose recognition sequence is contained in the primers. A kit can also contain several primers or several pairs of primers for use in genotyping at least two related or unrelated polymorphisms.

To carry out genotyping according to the invention, the primers are used to  
30 amplify a segment from a sample of template cDNA or genomic DNA. The term "amplification" as used herein refers to any process using a pair of primers

described above that produces multiple copies (ng amounts) of the segment of cDNA or genomic DNA between and including the portion complementary to the 5' ends of the pair of primers. The process of amplification can be carried out, for example, using the polymerase chain reaction (PCR) technique (*see, e.g.*, U.S. Patent 4,683,195 or reference 18) or by any other amplification method known in the art.

The amplified product can be cleaved using the restriction endonuclease whose recognition site is present in the primers. When the enzyme cleaves the DNA, it breaks a covalent bond at a discrete location on each strand. Digestion of a double stranded DNA molecule with a restriction endonuclease refers to the process of allowing the endonuclease to bind to its recognition site, cleave at its cleavage site, and release the cleaved DNA products. Because each member of the pair of primers of this invention contains a recognition site for the restriction endonuclease, digestion of the amplified product with the endonuclease will result in cleavage at two sites and consequently the release of a defined fragment or segment of the product. The product of the restriction endonuclease digestion will be a short, defined segment of double stranded DNA, whose length can be from 1, 2, 3, 4, 5, 6, 7, 8, 9, or 10 to about 16, 18, 20, 22, 24, 26, 30, 35, or 40 bp, but preferably is from about 7 to about 20 bp. The appropriate length of this segment is determined by the resolution of the MS method used for mass analysis. If the segment is too long, the analysis may be less sensitive.

The DNA cleavage product can be analyzed directly by ESI-MS or following an optional purification step. Purification can be carried out, for example, by reverse phase HPLC. The term "denature" refers to the dissociation of a double stranded DNA molecule to yield two single stranded DNA molecules. The "separation" of DNA molecules by ESI-MS refers to their physical separation from other molecules based on mass/charge ratio. The analysis can also be automated, for example, by performing the amplification and digestion steps in microtiter plates at a robotic workstation and loading the samples via an autosampler into an ESI-MS instrument. Loading on an HPLC can also be automated prior to ESI-MS. This would permit the rapid and sequential analysis

of a large number of polymorphic fragments, for example, obtained from a number of patients to be screened.

The above disclosure generally describes the present invention. A more complete understanding can be obtained by reference to the following specific examples, which are provided herein for purposes of illustration only and are not intended to limit the scope of the invention. Examples 2-4 present details of the ESI-MS analysis using polymorphisms of the human adenomatous polyposis carcinoma (APC) gene.

#### EXAMPLE 1

##### Generation of Short DNA Segments for ESI-MS

In order to unambiguously differentiate DNA fragments using a 2000 Da ion-trap mass spectrometer, it was first necessary to generate short, specific PCR products from complex genomes. To produce such short fragments (<20 bases), PCR amplification was carried out with primers containing a sequence for the type II restriction enzyme, *BpmI* (Figure 1).

Primers of 41-44 bases in length were designed so that 21 - 22 bases at the 5' end and 14 - 16 bases at the 3' end were precisely complementary to a 41 - 44 genomic sequence. The six base *BpmI* recognition sequence was placed between the 21-22 and 14-16 base portions, precisely replacing the 6 bases that were normally present at this position in the genome (Figure 1). Each PCR-primer contained at least 35 bases complementary to a specific genomic sequence, and the PCR fragments generated were only ~100 bp in length, thus ensuring that the PCR reaction was very robust.

PCR was performed as described (18). Reactions were performed with 25-50 ng of human genomic DNA, in 50 $\mu$ l. Thermal cycling conditions were 95°C for 2 min, followed by 40 cycles of 95°C for 30 sec., 60°C for 30 sec., and 72°C for 30 sec.

Following PCR amplification, low molecular weight oligonucleotides were obtained for mass analysis by restriction endonuclease digestion. 12  $\mu$ l of the PCR

product were digested with 10 units *BpmI* for 2 hours at 37°C in 50 µl. One unit of restriction endonuclease activity is the amount of enzyme required to completely digest 1 µg of substrate DNA in a 50 µl reaction in one hour at 37°C. DNA was extracted using one volume phenol/chloroform and precipitated in the presence of 3-5 µl of SeeDNA (Amersham), 6 volumes ethanol, and one third volume of 7.5 M ammonium acetate. After washing the pellets with 70% ethanol, the samples were allowed to air dry and resuspended in 10 µl of a solution of aqueous 0.4 M 1,1,1,3,3,3-hexafluoro-2-propanol (HFIP, Sigma) and methanol (85:15, v/v), of which 5 µl was typically injected for ESI-MS analysis.

After PCR amplification and *BpmI* digestion, DNA was purified by standard phenol/chloroform extraction and ethanol precipitation. It was not necessary to separate the larger (>40 bases) end fragments produced by *BpmI* digestion, as these were not confused with the short (7 - 20 bases), internal, variant fragments to be queried. Under ESI-MS conditions, these internal DNA fragments denatured and separated to produce detectable masses representing both the sense and antisense strands.

Oligonucleotide fragments for MS analysis were purified by reverse phase HPLC. Introduction of oligonucleotides into the HPLC coupled to the mass spectrometer was carried out at ~18 µl/min on a 15 cm x 800 µm ID Vydac C-18 reverse phase column (5 µm, 300 Å pore size, LC Packings, Amsterdam, NL). To obtain this flow rate, Waters 515 HPLC pumps (Waters Corp., Milford, MA, USA) operating at 0.2 ml/min were connected to an LC Packings Accurate microflow splitter. HPLC solvents were prepared from a stock solution of aqueous 0.8 M HFIP, adjusted to pH 7.0 with triethylamine, then diluted to 0.4 M (with water for solvent A and methanol for solvent B, as described by Apffel (19)). Initial analysis was carried out isocratically with a 20% A / 80% B solvent mixture (see, for example, Figure 2 and Figure 3). Alternatively, an initial solvent concentration of 70% A / 30% B was programmed to 50% A/50% B after 1 minute, where it was held for 10 minutes (see Figure 4 and Figure 5). The majority of potentially interfering compounds not removed by phenol/chloroform extraction and ethanol precipitation eluted with the void volume and were diverted

to waste, while oligonucleotides of interest were eluted as the methanol concentration was increased from 15% to 25%.

## EXAMPLE 2

### ESI-MS Analysis of the APC gene codon 1307 variant

5           This variant (I1307K) is present in 6% of Ashkenazi Jews, and is associated with a ~2-fold increase in colorectal cancer risk (7). The wild-type and variant sequences differ only at codon 1307 (ATA vs. AAA), and the A to T mutation represents the most difficult one to detect by MS analysis because the A to T change reflects only a 9 Da difference in mass.

10           Mass spectra were obtained on an LCQ ion-trap mass spectrometer (Finnigan MAT, San Jose, CA, USA) equipped with an electrospray ionization source operated in the negative ionization mode. To increase sensitivity, a 33 gauge stainless steel ESI needle, covered with 1/16" Teflon tubing outside the ESI source for insulation from the high voltage, was used in place of the standard  
15           fused silica ESI needle. The instrument was tuned daily by infusion at 1  $\mu$ l/min of one of the oligonucleotides studied (10 ng/ $\mu$ l in 70% A/30% B) into the 18  $\mu$ l/min HPLC mobile phase through a low dead-volume tee. Typical settings for the spray voltage were -2.5 to -5 kV. The stainless steel heated capillary temperature was held at 180°C.

20           Primers were designed according to the strategy outlined in Figure 1, so that 15-mer oligonucleotides were generated following *Bpm*I digestion. Primers used for PCR amplification of the APC variants were: 1307 sense (SEQ ID NO:1), 5'-AGACGACACAGGAAGCAGATTCTGGAGATACCCTGCAAATAGC-3; and 1307 antisense (SEQ ID NO:2),  
25           5'-GGAAGTTCGCTCACAGGATCTTCTGGAGACCTAGTTCCAATC-3'. The expected sizes of the product was 100 bp. Synthetically-generated antisense oligonucleotides, corresponding to two of the four expected fragments, were used to optimize the ESI-MS analysis. For both compounds, the most intense ions observed were  $[M-3H]^{3-}$  ions at m/z (mass to charge) 1519.3 (AAA) and m/z  
30           1522.3 (ATA) (Figure 2A and Figure 2B). The difference in m/z between these



[M-3H]<sup>3+</sup> ions is 3 Da, which was easily resolved under the experimental conditions (Figure 2C).

For detection of the I1307K variants (Figures 3A and 3B), the mass spectrometer was programmed to acquire data in the profile mode (1  $\mu$ scan; 1000 msec; isolation width 2.0 Da) using two scan events monitoring two [M-3H]<sup>3+</sup> ions simultaneously. Scan event 1: m/z 1581.7 [5'-pAGAAAAAAAAAGAAA-3', SEQ ID NO:3], 1519.3 [5'-pTTCTTTTTTTTCTGC-3', SEQ ID NO:4]. Scan event 2: m/z 1578.7 [5'-pAGAAATAAAAGAAA-3', SEQ ID NO:5], 1522.3 [5'-pTTCTTTTATTCTGC-3', SEQ ID NO:6]. Reconstructed ion chromatograms were generated and smoothed from this raw data using an isolation width of 1.0 Da and normalized to the largest of the four oligonucleotide ion peaks.

Genomic DNA was used as a template for PCR, and the PCR products digested with *BpmI* and purified by phenol/chloroform extraction. The samples were introduced into the mass spectrometer using the HPLC and [M-3H]<sup>3+</sup> ion masses characteristic of the two sense (m/z 1581.7 and 1587.7) and two antisense strands (m/z 1519.3 and 1522.3) were measured by selected ion monitoring as a function of time. It was found that there was sufficient material generated from the digestion of 1/4 of a 50  $\mu$ l PCR reaction for two ESI-MS injections. Furthermore, the simple phenol-chloroform purification was sufficient to obtain good mass chromatographic peaks with minimal interference from other compounds in the channels monitored (Figure 3). Sixteen human samples, which had previously been analysed by sequencing, were genotyped with this method. Samples from subjects who were heterozygous had peaks in all four channels monitored (*i.e.*, had both the wild-type and mutant sense and antisense strands), whereas samples from individuals who were homozygous for the wild-type allele only had peaks in the two wild-type channels. There was 100% concordance between SOMA and sequencing results.

### EXAMPLE 3

#### ESI-MS Analysis of the APC Codon 1493 Variant

A second variant in the APC gene (ACA or ACG at codon 1493) was



selected to demonstrate the general applicability of the methodology, even in difficult cases. This variant is not associated with disease, but is a common polymorphism which can be used for linkage analysis in families with familial adenomatous polyposis (8).

5           Primers used for PCR amplification of the APC variants were: 1493 sense ,  
5'-TTCAGAGGGTCCAGGTTCTTCCTGGAGCTGATACTTTATTACA-3' (SEQ ID  
NO:7); and 1493 antisense,  
5'GCACTCAGGCTGGATGAACAACCTGGAGCCATCTGGAGTACT-3' (SEQ ID  
NO:8). The expected size of the product was 100 bp. The internal fragments  
10           generated by SOMA were designed to be 16 bp long. Moreover, for one of the  
alleles (ACG), the sense (5'-pTTTTGCCACGGAAAGT-3', SEQ ID NO:9) and  
antisense (5'-pTTTCCGTGGCAAAATG-3', SEQ ID NO:10) oligonucleotides had  
different base sequences but the same mass. This resulted in two oligonucleotide  
15           [M-3H]<sup>2+</sup> ions with identical mass-to-charge ratios at 1657.7 which could not be  
resolved by ESI-MS. However, it was found that ESI-MS/MS selected reaction  
monitoring could easily differentiate between the four oligonucleotide ions.  
Heterozygotes were identified by the presence of chromatographic peaks in all  
four channels, while peaks in the sense and antisense channels of one allele  
indicated a homozygous sample (Figure 4). Of 50 individuals genotyped at codon  
20           1493, there was a 100% correlation between the results obtained by SOMA and  
sequencing.

          Although it might be expected that the four chromatographic peaks obtained  
for the four oligonucleotides produced from a heterozygote would be of equal  
intensity, this is not always the case. Oligonucleotide base sequence, length, and  
25           conformation cause variations in ESI-MS response factors. However, for all  
variants we studied, the relative response factors measured for synthetic  
oligonucleotide standards closely approximated those measured for the four  
oligonucleotides generated from human DNA. This allowed straightforward  
normalization of the signals obtained if desired, though no normalization was used  
30           in the data presented in Figures 2 - 5. Interference from oligonucleotide-cation  
adducts and non-specific DNA fragments produced background signals for certain

variants. This background could be reduced with improved chromatographic separation and sample cleanup, or by simply redesigning the primers for amplification to produce a slightly different internal fragment containing the sequence variation. MS/MS is also a powerful technique for improving selectivity, even in the presence of interfering compounds (Figure 4). To date, SOMA has been used to analyze seven different single nucleotide variations. Of these, all four species (sense and antisense from the two alleles) could be readily discerned in five cases on the first try, while in two cases, different primers, producing a slightly different length or position of interrogated sequence, had to be designed to produce acceptable results.

#### EXAMPLE 4

##### Simultaneous Analysis of Multiple Variants

Three common polymorphisms in the APC gene, at codons 485, 545, and 1756 (8), were chosen to demonstrate that multiple polymorphisms could be analysed in parallel by ESI-MS.

For detection of multiple variants (Figure 5), the mass spectrometer was programmed to acquire data in the profile mode (1  $\mu$ scan; 30 msec; isolation width 3.0 Da) using two  $\sim$ 1.4-sec scan events monitoring 16  $[M-2H]^{2-}$  ions simultaneously. (Scan event 1: 486-TAC-s  $m/z$  1271.8 [5'-pTGTACGGG-3']; 486-TAC-as  $m/z$  1231.3 [5'-pCGTACATT-3']; 545-GCA-s  $m/z$  1407.9 [5'-pATTGCAAGT-3']; 545-GCA-as  $m/z$  1399.9 [5'-pTTGCAATAA-3']; 1756-TCG-s  $m/z$  1688.6 [5'-pGCGTCGTCTTC-3', SEQ ID NO:11]; 1756-TCG-as  $m/z$  1726.6 [5'-pAGACGACGCAG-3', SEQ ID NO:12]. Scan event 2: 486-TAT-s  $m/z$  1279.3 [5'-pTGTATGGG-3']; 486-TAT-as  $m/z$  1223.3 [5'-pCATAACATT-3']; 545-GCG-s  $m/z$  1415.9 [5'-pATTGCGAGT-3']; 545-GCG-as  $m/z$  1392.4 [5'-pTCGCAATAA-3']; 1756-TCT-s  $m/z$  1676.1 [5'-pGCGTCTTCTTC3', SEQ ID NO:13]; 1756-TCT-as  $m/z$  1738.6 [5'-pAGAAGACGCAG-3', SEQ ID NO:14]). Reconstructed ion chromatograms were generated and smoothed from this raw data using an isolation width of 1.0 Da and normalized to the largest of the four oligonucleotide ion peaks for each

variant.

For analysis of DNA segments that have identical masses but different nucleotide sequences, the technique of tandem MS (MS/MS) was applied to distinguish the segments. ESI-MS/MS was used for analysis of the 1493 variant (Figure 4). Using this technique, the four oligonucleotide ions studied were isolated in the ion-trap and subjected to collisional-induced dissociation at 60% collision energy, resulting in sequence-specific fragment ions of the four original ions. Signals from two MS/MS fragment ions were summed as a function of time for each of the four oligonucleotide  $[M-3H]^3$  ions monitored. The mass spectrometer was programmed to acquire data in the profile mode (1  $\mu$ scan; 500 msec; isolation width 3.5 Da) using four scan events monitoring each  $[M-2H]^2$  oligonucleotide ion individually. (Scan event 1: ACG-s: m/z 1657.7  $\rightarrow$  1392.9+1589.0. Scan event 2: ACG-as: m/z 1657.7  $\rightarrow$  1089.1+1667.1. Scan event 3: ACA-s: m/z 1652.4  $\rightarrow$  1393.1+1589.2. Scan event 4: ACA-as: m/z 1662.7  $\rightarrow$  1089.1+1682.0.) Reconstructed ion chromatograms were generated and smoothed from this raw data using an isolation width of 1.0 Da and normalized to the largest of the four oligonucleotide ion peaks.

Primers used for PCR amplification of the variants were: 486 sense, 5'-GGACTACAGGCCATTGCAGAACTGGAGCAAGTGGACTGTGAAA-3' (SEQ ID NO:15); 486 antisense, 5'-AGCATATCGTCTTAGTGTAATACTGGAGTGGTCATTAGTAAG-3' (SEQ ID NO:16); 545 sense, 5'-ATTTTATGTATAAAATTAATCTCTGGAGGATTAATTTGCAGGTT-3' (SEQ ID NO:17); 545 antisense, 5'-TTTACTATTTACATCTGCTCGCCTGGAGAAATTCCTCAAAAC-3' (SEQ ID NO:18); 1756 sense, 5'-TTTCCGTGTGAAAAAGATAATCTGGAGGGTCCAGCAAGCATCT-3' (SEQ ID NO:19); and 1756 antisense, 5'-GGTTTCTTTTCTTACCATCTACTGGAGTTTGTGTTGGGTGCA-3' (SEQ ID NO:20). The expected sizes of the products were 93 bp for codon 486, 94bp for codon 545, and 96bp for codon 1756. Regions around each of the polymorphic

5 sites were amplified in separate PCR reactions and *Bpm*I digestion was performed, producing DNA fragments of 8, 9, and 11 bases containing codons 485, 545, and 1756, respectively. The three reaction mixtures from each individual were then combined, purified by phenol-chloroform extraction, and introduced into the mass spectrometer using the HPLC. Twelve  $[M-2H]^{2-}$  ion masses, characteristic of the three polymorphisms, were monitored by ESI-MS selected ion monitoring. Results for simultaneous determination of polymorphisms at the three codons in two individuals homozygous for each polymorphism are shown in Figure 5. Heterozygotes displayed the expected four peaks (not shown). The results  
10 obtained by SOMA and sequencing were again perfectly concordant.

Prior methods of identifying sequence variations in human DNA by MS have for the most part employed matrix-assisted laser desorption/ionisation time-of-flight mass spectrometry (MALDI-TOF). With that technique, a UV laser pulse to the sample in a fixed matrix causes ionized biomolecules to be released  
15 into the gas phase where they can be extracted for mass separation. MALDI-TOF has been used most successfully to analyze variations which are characterized by large mass differences (5, 11, 12). When used to identify SNPs, the use of MALDI-TOF has usually required hybridization of small fragments to PCR-amplified DNA for adequate resolution (6, 13-16). In addition, use of the  
20 technique has been hampered by interference from sodium and potassium adduct ions, which can lead to errors in the determination of ion mass and decreased signal-to-noise ratios.

Although PCR has previously been coupled with ESI-MS to assess insertion/deletion-type variations in human DNA (17), this invention represents  
25 the first application of ESI-MS to detect SNPs. The ESI mass spectrum gives information on both alleles and for both sense and antisense strands. The approach is applicable to any subtle variation and can measure the variations with the smallest possible mass difference with excellent resolution. The method requires just picomole quantities of oligonucleotide for each analysis. Sample  
30 clean-up, involving standard phenol/chloroform extraction and ethanol

precipitation, is simple, quick and amenable to automation.

### **References**

1. Struewing, J.P., Hartge, P., Wacholder, S., Baker, S.M., Berlin, M.,  
McAdams, M., et al. 1997. The risk of cancer associated with specific  
5 mutations of BRCA1 and BRCA2 among Ashkenazi Jews. *N. Engl. J. Med.*  
336:1401-1408
2. Martinez, M., Campion, D., Brice, A., Hannequin, D., Dubois, B., Didierjean,  
O., et al. 1998. Apolipoprotein E epsilon4 allele and familial aggregation of  
Alzheimer disease. *Arch. Neurol.* 55:810-816.
- 10 3. De Stefano, V., Chiusolo, P., Paciaroni, K., Casorelli, I., Rossi, E., Molinari,  
M. et al. 1998. Prothrombin G20210A mutant genotype is a risk factor for  
cerebrovascular ischemic disease in young patients. *Blood* 91:3562-3565.
4. Nollau, P. and Wagener, C. 1997. Methods for detection of point mutations:  
performance and quality assessment. IFCC Scientific Division, Committee on  
15 Molecular Biology Techniques. *Clin. Chem.* 43:1114-1128.
5. Ross, P.L. and Belgrader, P. 1997. Analysis of short tandem repeat  
polymorphisms in human DNA by matrix- assisted laser desorption/ionization  
mass spectrometry. *Anal. Chem.* 69:3966-3972.
6. Griffin, T.J., Tang, W. and Smith, L.M. 1997. Genetic analysis by peptide  
20 nucleic acid affinity MALDI-TOF mass spectrometry. *Nat. Biotechnol.*  
15:1368-1372.
7. Laken, S. J., Petersen, G. M., Gruber, S. B., Oddoux, C., Ostrer, H.,  
Giardiello, F. M. et al. 1997. Familial colorectal cancer in Ashkenazim due to  
a hypermutable tract in APC. *Nat. Genet.* 17:79-83.
- 25 8. Powell, S. M., Zilz, N., Beazer-Barclay, Y., Bryan, T. M., Hamilton, S. R.,  
Thibodeau, S. N. et al. 1992. APC mutations occur early during colorectal  
tumorigenesis. *Nature* 359:235-237.
9. Miletova, P. and Schram, K.H. 1997. Mass spectrometry of nucleotides and  
oligonucleotides. *Mol. Biotechnol.* 8:249-253.
- 30 10. Crain, P.F. and McCloskey, J.A. 1998. Applications of mass spectrometry to

the characterization of oligonucleotides and nucleic acids. *Curr. Opin. Biotechnol.* 9:25-34.

11. Braun, A., Little, D.P., Reuter, D., Muller-Mysok, B. and Koster, H. 1997. Improved analysis of microsatellites using mass spectrometry. *Genomics* 46:18-23.
12. Wada, Y. 1998. Separate analysis of complementary strands of restriction enzyme- digested DNA. An application of restriction fragment mass mapping by matrix-assisted laser desorption/ionization mass spectrometry. *J. Mass. Spectrom.* 33:187-192.
13. Ross, P.L., Lee, K. and Belgrader, P. 1997. Discrimination of single-nucleotide polymorphisms in human DNA using peptide nucleic acid probes detected by MALDI-TOF mass spectrometry. *Anal. Chem.* 69:4197-202.
14. Fei, Z., Ono, T. and Smith, L.M. 1998. MALDI-TOF mass spectrometric typing of single nucleotide polymorphisms with mass-tagged ddNTPs. *Nucleic Acids Res.* 26:2827-2828.
15. Higgins, G.S., Little, D.P. and Koster, H. 1997. Competitive oligonucleotide single-base extension combined with mass spectrometric detection for mutation screening. *Biotechniques* 23:710-714.
16. Haff, L.A. and Smirnov, I.P. 1997. Single-nucleotide polymorphism identification assays using a thermostable DNA polymerase and delayed extraction MALDI-TOF mass spectrometry. *Genome Res.* 7:378-388.
17. Naito, Y., Ishikawa, K., Koga, Y., Tsuneyoshi, T., Terunuma, H., and Arakawa, R. 1997. Genetic Diagnosis by Polymerase Chain Reaction and Electrospray Ionization Mass Spectrometry: Detection of Five Base Deletion From Blood DNA of a Familial Adenomatous Polyposis Patient. *J. Am. Soc. Mass Spectrom.* 8:737-742.
18. Riggins, G.J., Kinzler, K.W., Vogelstein, B. and Thiagalingam, S. 1997. Frequency of Smad gene mutations in human cancers. *Cancer Res.* 57:2578-2580.
19. Apffel, A., Chakel, J.A., Fischer, S., Lichtenwalter, K. and Hancock, W.S.

1997. Analysis of Oligonucleotides by HPLC-Electrospray Ionization Mass Spectrometry. *Anal. Chem.* 69:1320-1325.
20. Szybalski, W. 1985. Universal Restriction Endonucleases: Designing Novel Cleavage Specificities by Combining Adapter Oligodeoxynucleotide and Enzyme Moieties. *Gene* 40:169-173.
21. Friedman, S.H. and Kenyon, G.L. 1995. Bioorganic Chemistry. In *Molecular Biology and Biotechnology: A Comprehensive Desk Reference*, R.A. Meyers, Ed., Wiley-VCH, NY, p. 103-106.

**WE CLAIM:**

1. An isolated primer for amplifying a segment of DNA comprising:  
a linear oligonucleotide comprising a 5' end and a 3' end, said  
oligonucleotide consisting of at least 35 nucleotides, wherein a first portion  
of said oligonucleotide of at least 13 nucleotides at the 5' end of said  
oligonucleotide and a second portion of the oligonucleotide of from 5 to 22  
nucleotides at the 3' end of the oligonucleotide are precisely complementary  
to a first portion and a second portion, respectively, of a segment of a  
cDNA or genomic DNA, wherein 4-8 nucleotides between the first portion  
and the second portion of the oligonucleotide comprise a recognition site for  
a restriction endonuclease that cleaves at least 5 nucleotides from its  
recognition site, wherein the segment of the cDNA or genomic DNA does  
not comprise the recognition site.
2. The primer of claim 1 wherein the segment of the cDNA or genomic DNA  
to which the first and second portions of the oligonucleotide are  
complementary comprises from 0 to 12 nucleotides between the first portion  
of said segment and the second portion of said segment.
3. The primer of claim 1 wherein the segment of the cDNA or genomic DNA  
to which the first and second portions of the oligonucleotide are  
complementary comprises from 4 to 8 nucleotides between the first portion  
of said segment and the second portion of said segment.
4. The primer of claim 1 wherein the segment of the cDNA or genomic DNA  
to which the first and second portions of the oligonucleotide are  
complementary comprises 6 nucleotides between the first portion of said  
segment and the second portion of said segment.
5. The primer of claim 1 wherein the restriction endonuclease is a Type IIS  
restriction endonuclease.



6. The primer of claim 5 wherein the Type IIS restriction endonuclease is BpmI.
7. The primer of claim 1 wherein the restriction endonuclease cleaves DNA at least 8 nucleotides from its recognition site.
- 5 8. An isolated primer for amplifying a segment of DNA comprising:  
a linear oligonucleotide comprising a 5' end and a 3' end, said  
oligonucleotide consisting of at least 35 nucleotides, wherein a first portion  
of said oligonucleotide of at least 13 nucleotides at the 5' end of said  
oligonucleotide and a second portion of the oligonucleotide of from 5 to 22  
10 nucleotides at the 3' end of the oligonucleotide are substantially  
complementary to a first portion and a second portion, respectively, of a  
segment of a cDNA or genomic DNA, wherein 4-8 nucleotides between the  
first portion and the second portion of the oligonucleotide comprise a  
15 recognition site for a restriction endonuclease that cleaves at least 5  
nucleotides from its recognition site, wherein the segment of the cDNA or  
genomic DNA does not comprise the recognition site.
9. The primer of claim 8 wherein the segment of the cDNA or genomic DNA  
to which the first and second portions of the oligonucleotide are  
complementary comprises from 0 to 12 nucleotides between the first portion  
20 of said segment and the second portion of said segment.
10. The primer of claim 8 wherein the segment of the cDNA or genomic DNA  
to which the first and second portions of the oligonucleotide are  
complementary comprises from 4 to 8 nucleotides between the first portion  
of said segment and the second portion of said segment.
- 25 11. The primer of claim 8 wherein the segment of the cDNA or genomic DNA

to which the first and second portions of the oligonucleotide are complementary comprises 6 nucleotides between the first portion of said segment and the second portion of said segment.

- 5 12. The primer of claim 8 wherein the restriction endonuclease is a Type IIS restriction endonuclease.
13. The primer of claim 12 wherein the Type IIS restriction endonuclease is BpmI.
14. The primer of claim 8 wherein the restriction endonuclease cleaves DNA at least 8 nucleotides from its recognition site.
- 10 15. An isolated pair of primers for amplifying a segment of cDNA or genomic DNA,  
wherein each primer comprises a linear oligonucleotide comprising a 5' end and a 3' end, said oligonucleotide consisting of at least 35 nucleotides, wherein a first portion of said oligonucleotide of at least 13 nucleotides at  
15 the 5' end of said oligonucleotide and a second portion of the oligonucleotide of from 5 to 22 nucleotides at the 3' end of the oligonucleotide are precisely complementary to a first portion and a second portion of a cDNA or genomic DNA, wherein 4-8 nucleotides between the first portion and the second portion of the oligonucleotide comprise a  
20 recognition site for a restriction endonuclease that cleaves at least 5 nucleotides from its recognition site, wherein the segment of the cDNA or genomic DNA does not comprise the recognition site for the restriction endonuclease,  
wherein each primer of the pair of primers is complementary to an  
25 opposite strand of a double stranded cDNA or genomic DNA molecule, wherein the pair of primers is complementary to two non-contiguous portions of the double stranded cDNA or genomic DNA molecule, wherein

1 to 20 nucleotides separate the two non-contiguous portions of the double stranded cDNA or genomic DNA molecule.

16. The pair of primers of claim 15 wherein the restriction endonuclease is a Type IIS restriction endonuclease.
- 5 17. The pair of primers of claim 15 wherein a single nucleotide polymorphism maps to the 1 to 20 nucleotides which separate the two non-contiguous portions of the double stranded DNA molecule.
18. The pair of primers of claim 15 which are contained in a kit.
- 10 19. The pair of primers of claim 18 wherein the kit further comprises the restriction endonuclease.
20. A kit comprising a plurality of pairs of primers according to claim 15.
21. An isolated pair of primers for amplifying a segment of cDNA or genomic DNA,  
wherein each primer comprises a linear oligonucleotide comprising a  
15 5' end and a 3' end, said oligonucleotide consisting of at least 35 nucleotides,  
wherein a first portion of said oligonucleotide of at least 13 nucleotides at  
the 5' end of said oligonucleotide and a second portion of the  
oligonucleotide of from 5 to 22 nucleotides at the 3' end of the  
oligonucleotide are substantially complementary to a first portion and a  
20 second portion of a cDNA or genomic DNA, wherein 4-8 nucleotides  
between the first portion and the second portion of the oligonucleotide  
comprise a recognition site for a restriction endonuclease that cleaves at  
least 5 nucleotides from its recognition site, wherein the segment of the  
cDNA or genomic DNA does not comprise the recognition site for the  
25 restriction endonuclease,

- 5                    wherein each primer of the pair of primers is complementary to an  
opposite strand of a double stranded cDNA or genomic DNA molecule,  
wherein the pair of primers is complementary to two non-contiguous  
portions of the double stranded cDNA or genomic DNA molecule, wherein  
1 to 20 nucleotides separate the two non-contiguous portions of the double  
stranded cDNA or genomic DNA molecule.
22.   The pair of primers of claim 21 wherein the restriction endonuclease is a  
Type IIS restriction endonuclease.
- 10   23.   The pair of primers of claim 21 wherein a single nucleotide polymorphism  
maps to the 1 to 20 nucleotides which separate the two non-contiguous  
portions of the double stranded DNA molecule.
24.   The pair of primers of claim 21 which are contained in a kit.
25.   The pair of primers of claim 21 wherein the kit further comprises the  
restriction endonuclease.
- 15   26.   A kit comprising a plurality of pairs of primers according to claim 21.
27.   A method for producing a short segment of DNA, suitable for analysis by  
mass spectrometry, comprising the steps of:  
                                 amplifying cDNA or genomic DNA of a subject using the pair of  
primers of claim 15 to form amplified DNA;  
20                                digesting the amplified DNA with the restriction endonuclease to  
form a short segment of DNA.
28.   A method for producing a short segment of DNA, suitable for analysis by  
mass spectrometry, comprising the steps of:  
                                 amplifying cDNA or genomic DNA of a subject using the pair of

primers of claim 21 to form amplified DNA;

digesting the amplified DNA with the restriction endonuclease to form a short segment of DNA.

- 5 29. A method for analyzing a first short segment of DNA comprising a first polymorphic nucleotide to distinguish the first short segment of DNA from a second short segment of DNA comprising a second polymorphic nucleotide, the method comprising the steps of:

10 applying a mixture of DNA segments to an electrospray ionization/mass spectrometer, whereby the DNA segments are denatured and the denatured segments are separated, wherein the mixture of DNA segments is made by the process of:

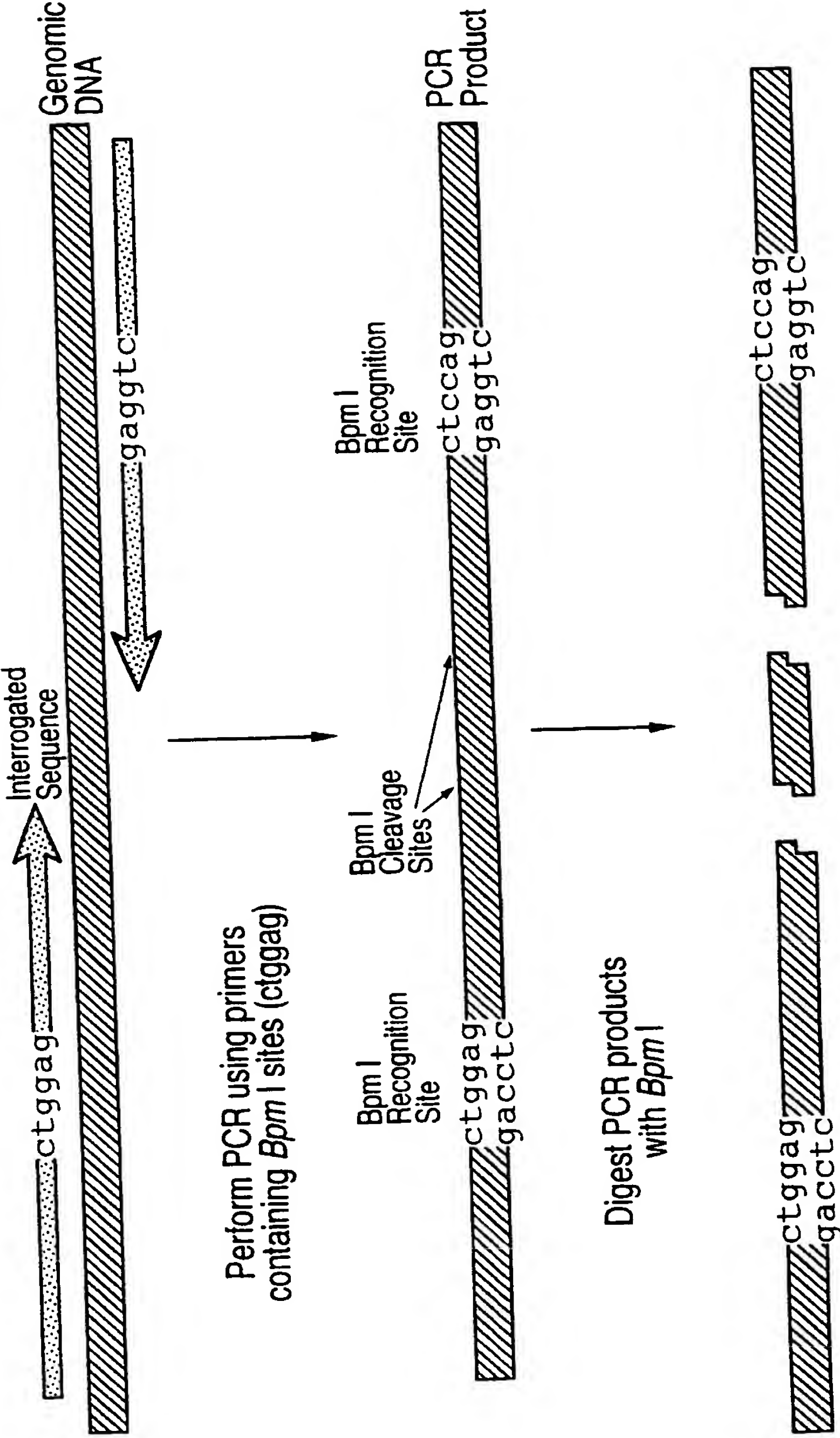
15 amplifying cDNA or genomic DNA of a subject using the pair of primers of claim 15 to form amplified DNA; and digesting the amplified DNA with the restriction endonuclease to form a short segment of DNA.

30. A method for analyzing a first short segment of DNA comprising a first polymorphic nucleotide to distinguish the first short segment of DNA from a second short segment of DNA comprising a second polymorphic nucleotide, the method comprising the steps of:

20 applying a mixture of DNA segments to an electrospray ionization/mass spectrometer, whereby the DNA segments are denatured and the denatured segments are separated, wherein the mixture of DNA segments is made by the process of:

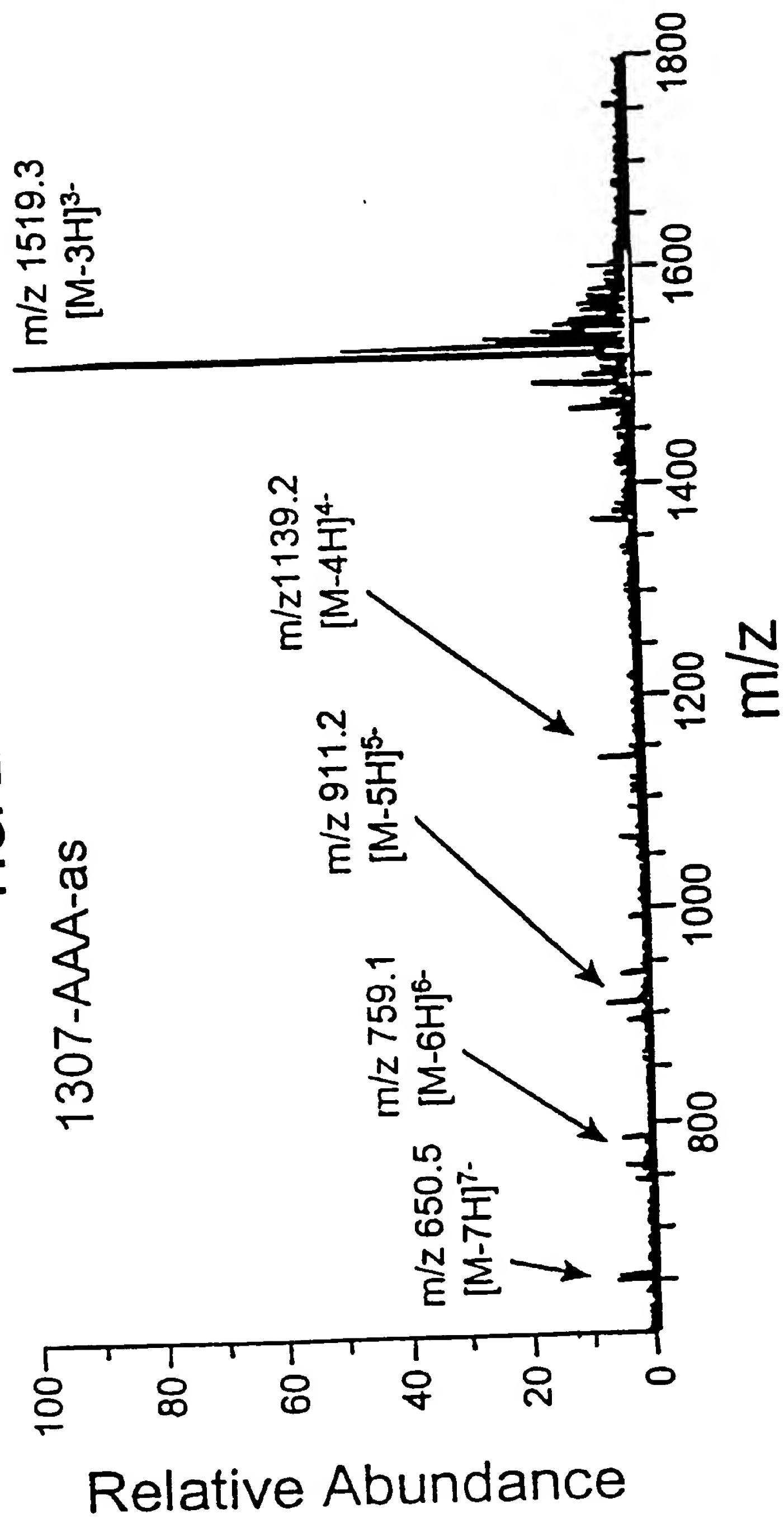
25 amplifying cDNA or genomic DNA of a subject using the pair of primers of claim 21 to form amplified DNA; and digesting the amplified DNA with the restriction endonuclease to form a short segment of DNA.

FIG. 1



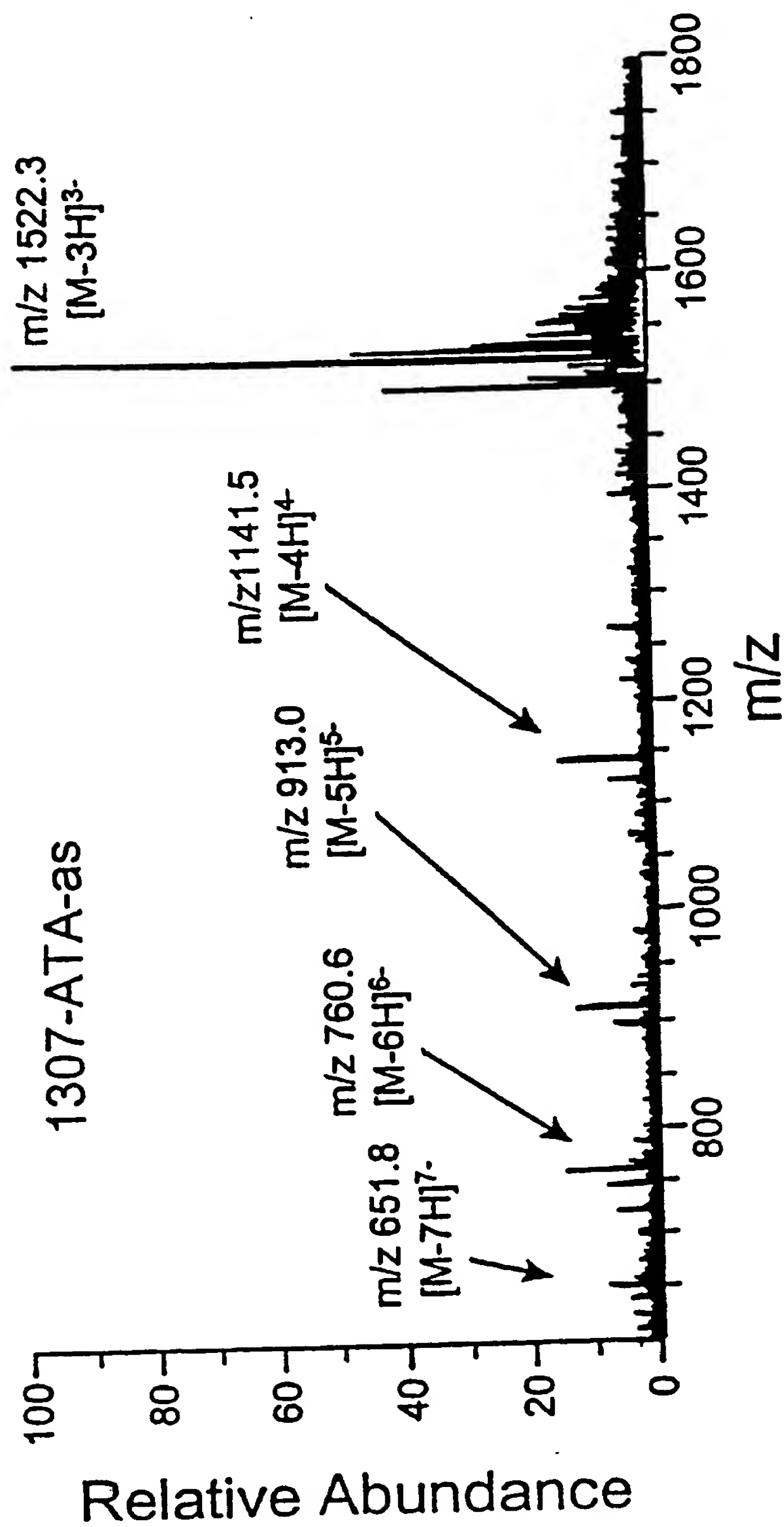
2/11

FIG. 2A



3/11

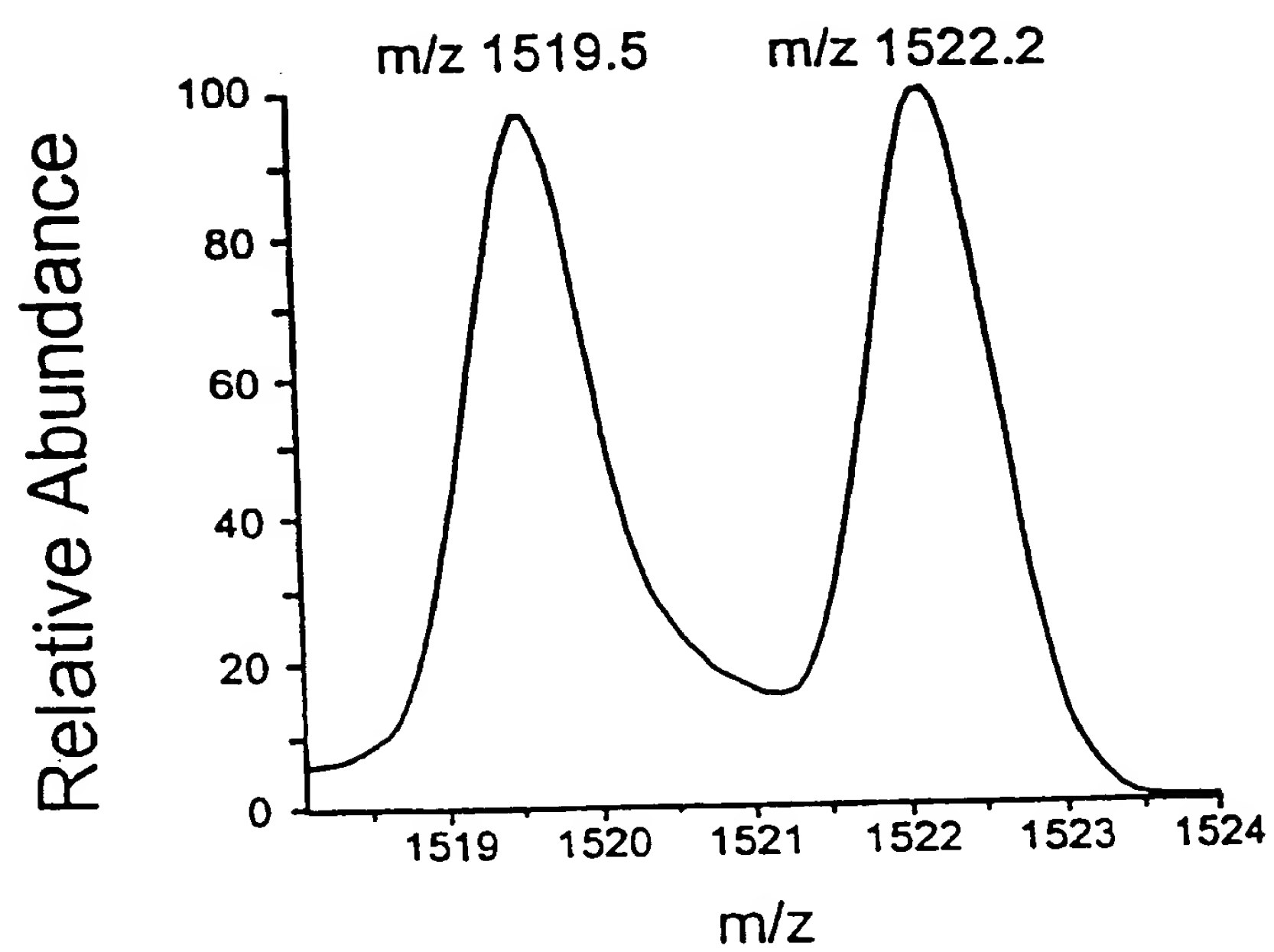
FIG. 2B



SUBSTITUTE SHEET (RULE 26)

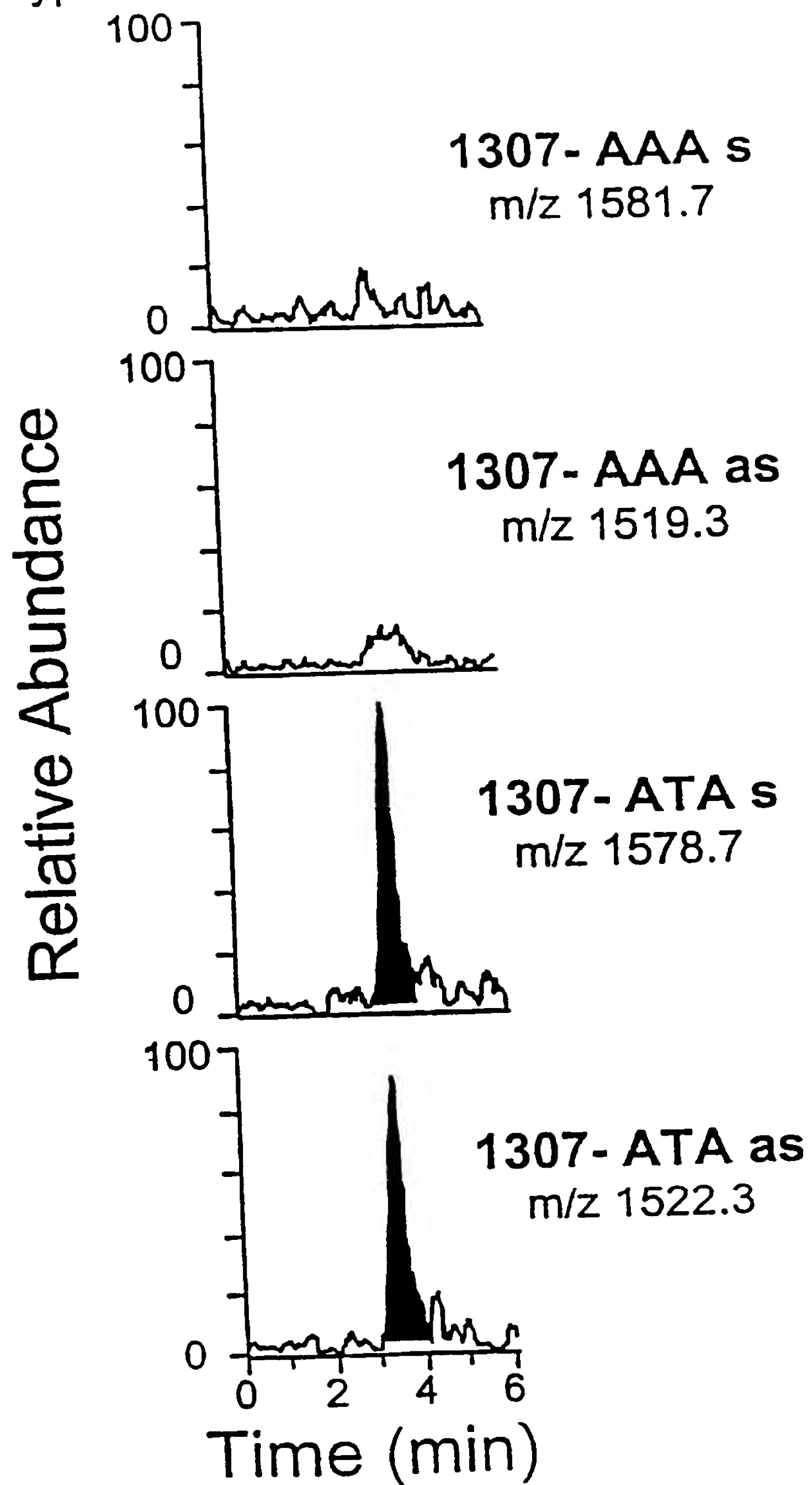


4/11

**FIG. 2C**

**FIG. 3A**<sup>5/11</sup>

Genotype: ATA/ATA



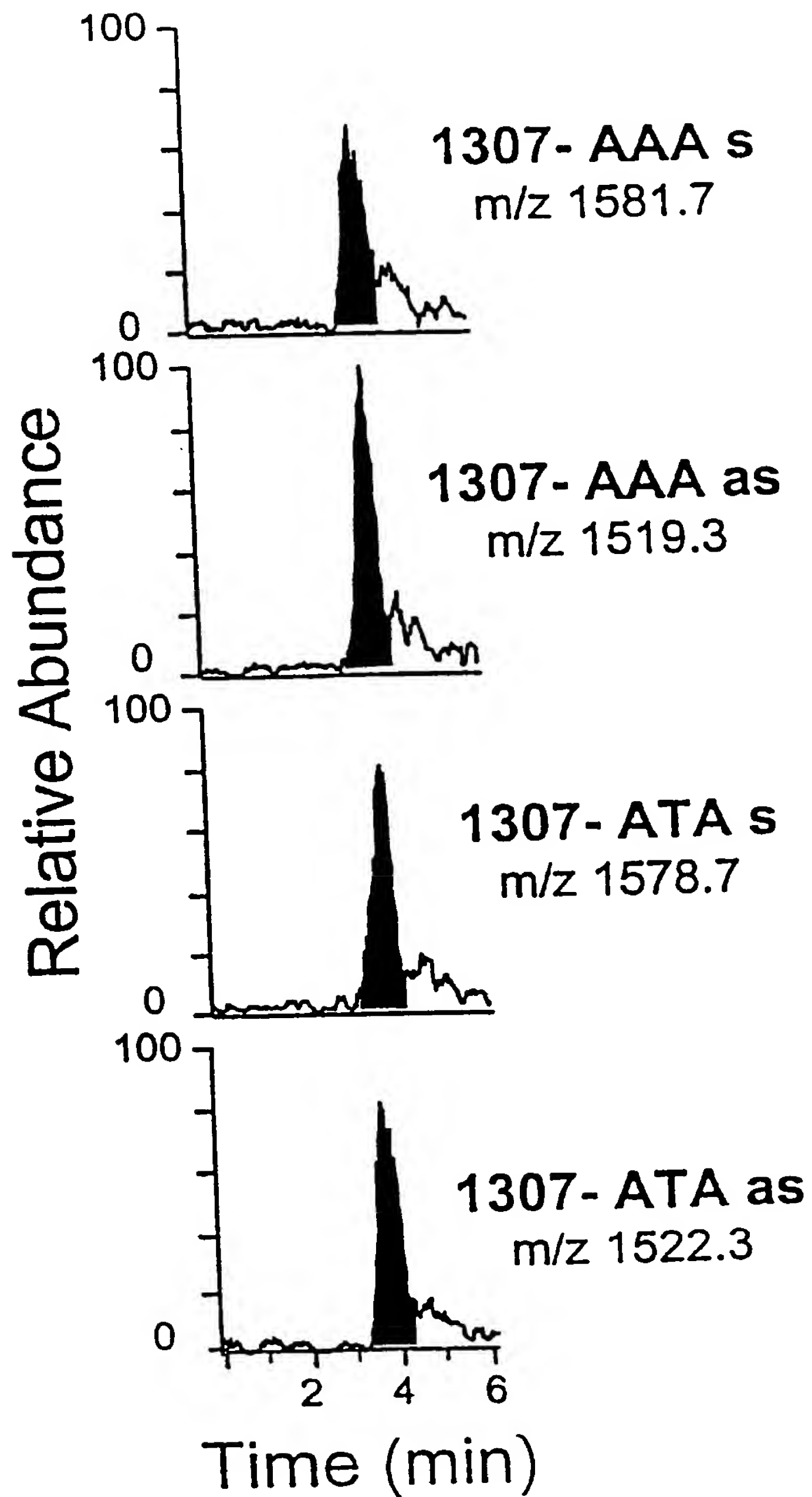
SUBSTITUTE SHEET (RULE 26)

6/11

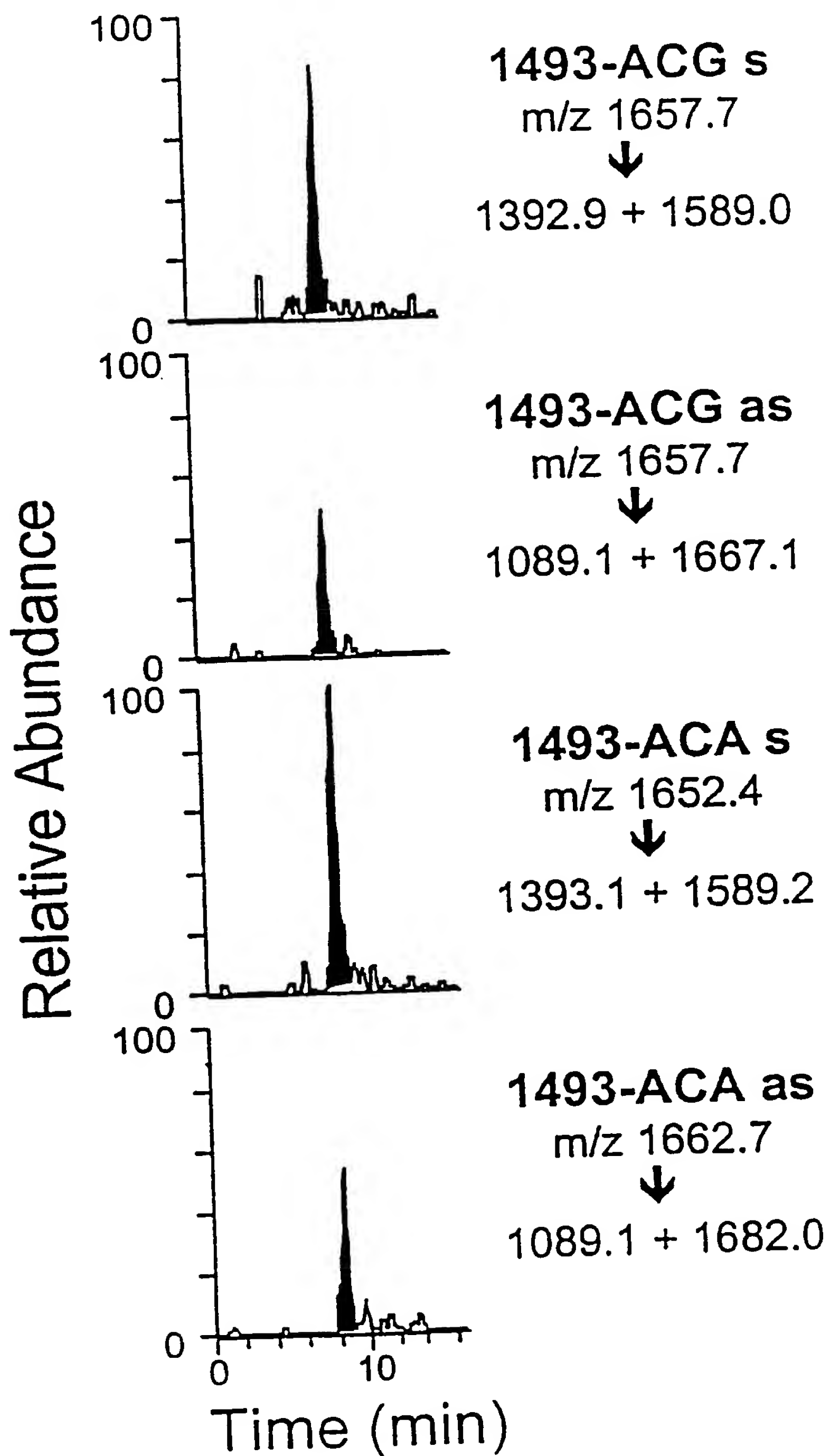
**FIG. 3B**

Genotype:

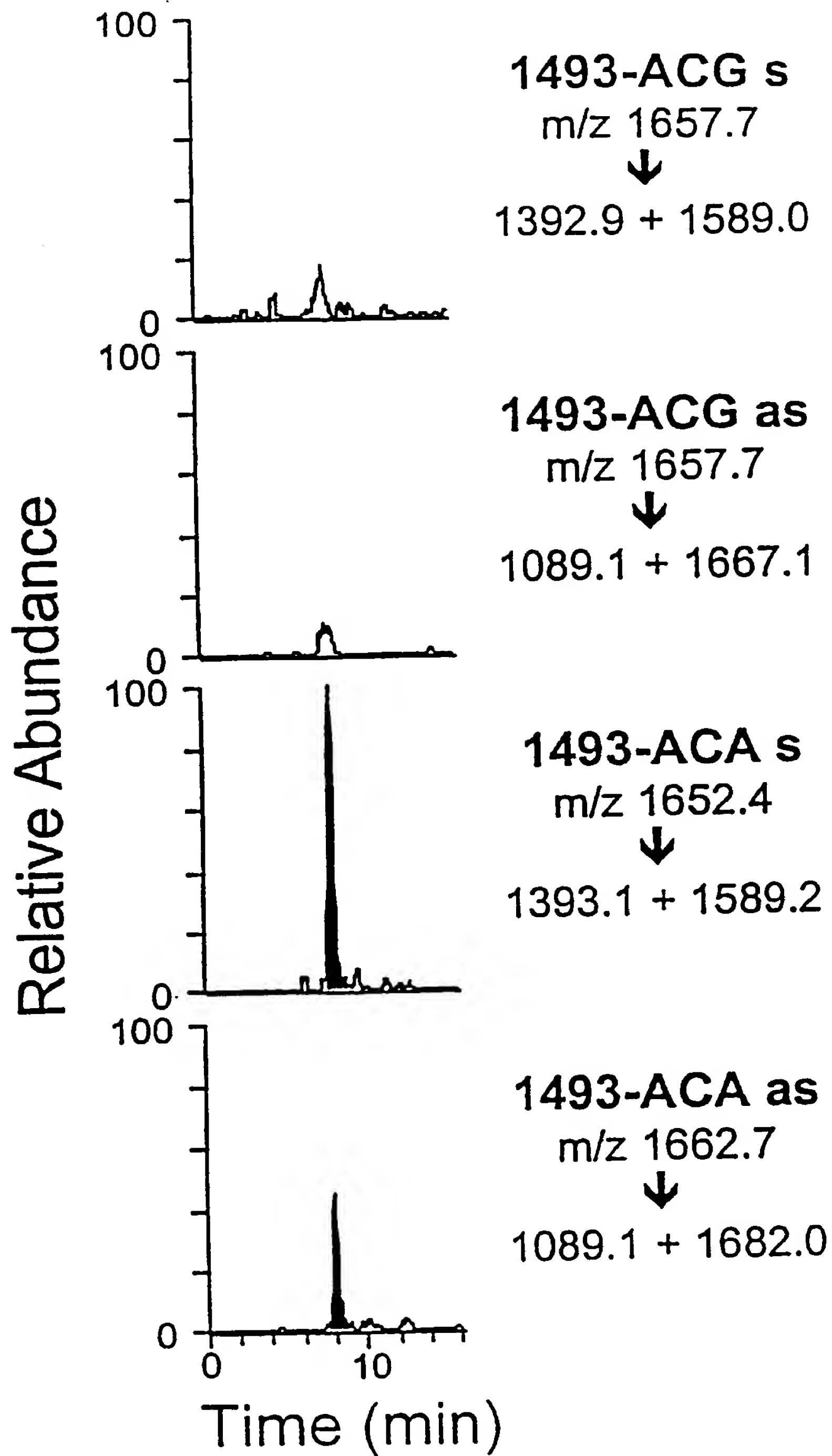
ATA/AAA

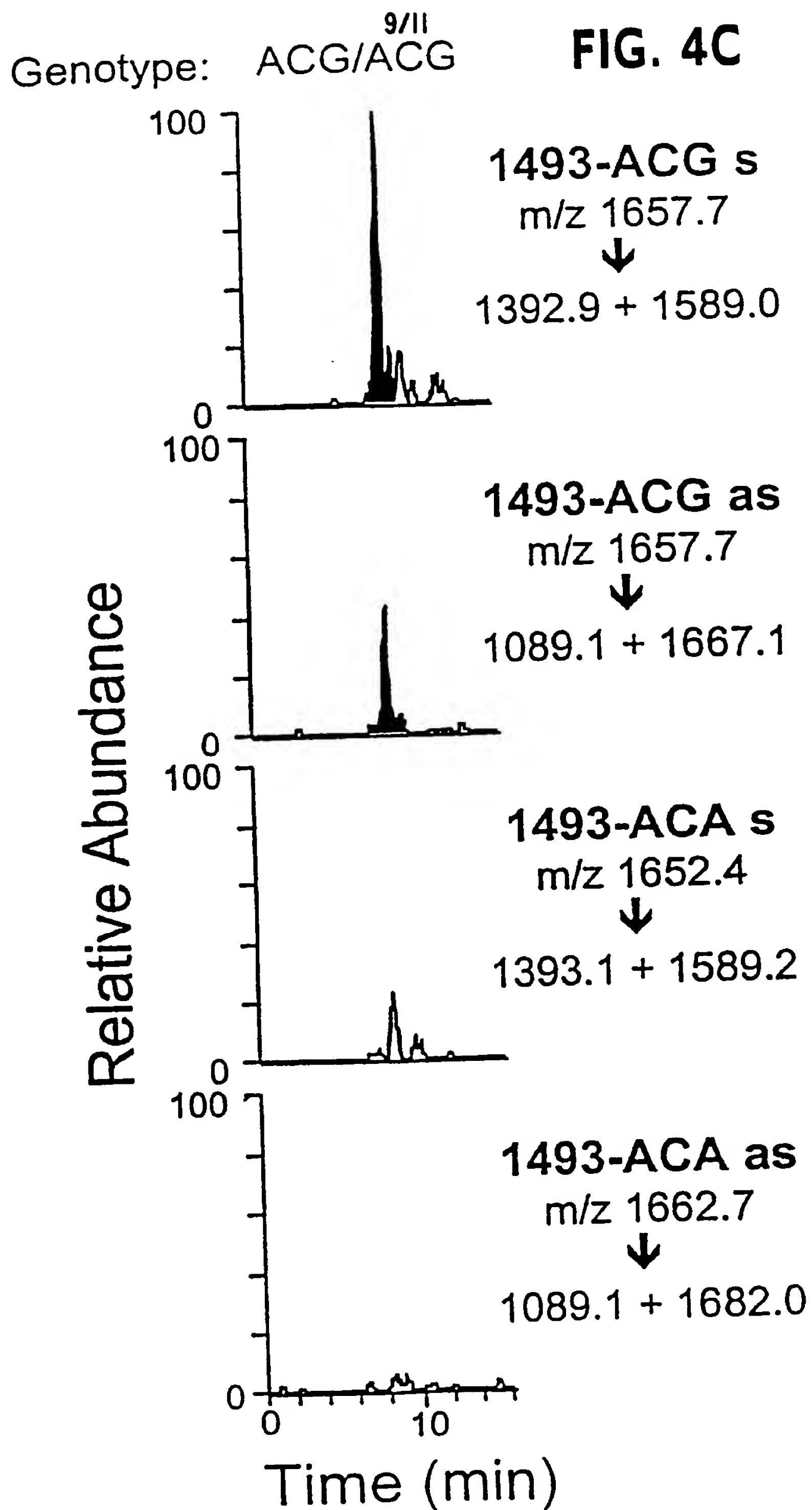


SUBSTITUTE SHEET (RULE 26)

Genotype: ACG/ACA<sup>7/11</sup>**FIG. 4A**

SUBSTITUTE SHEET (RULE 26)

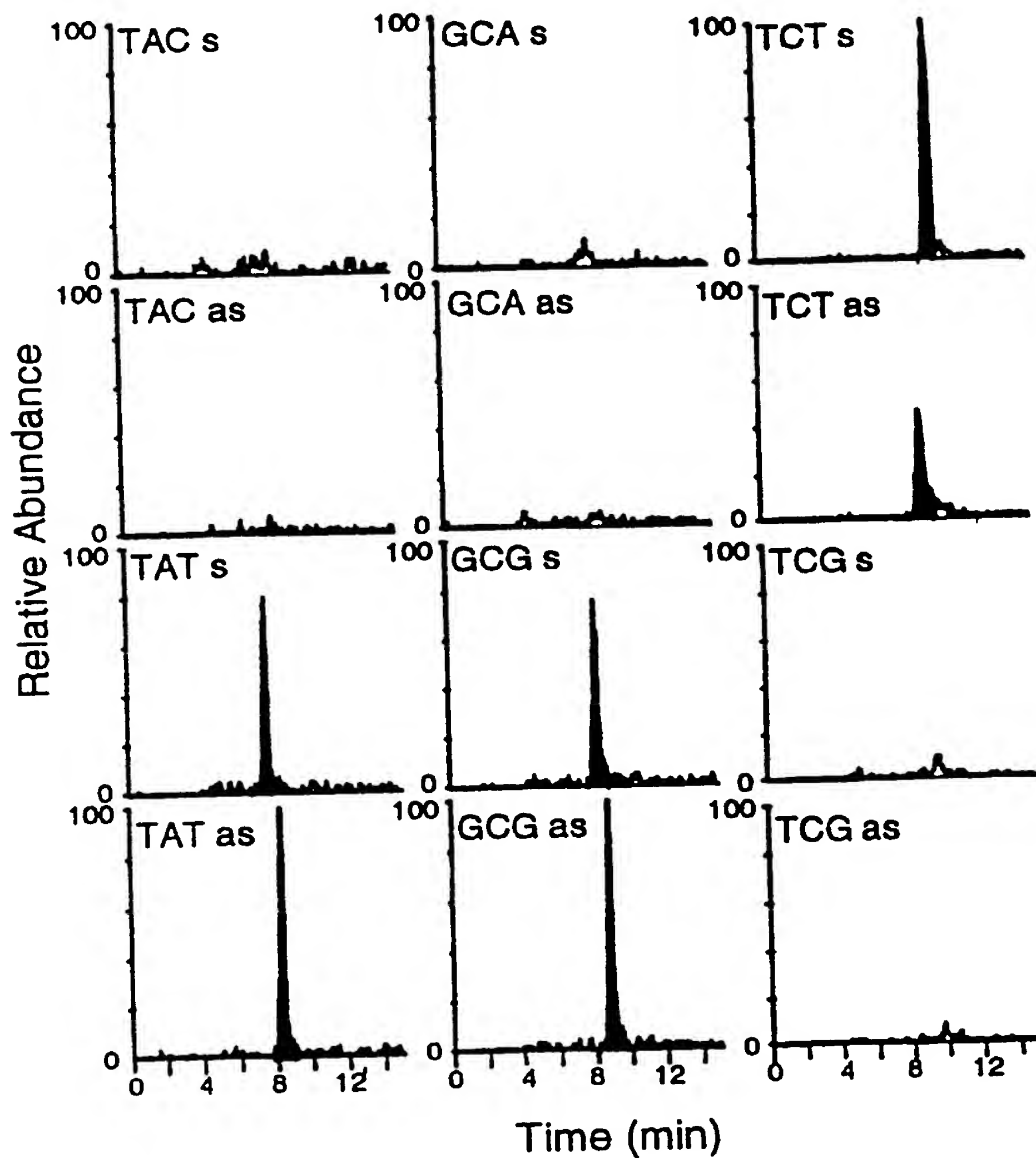
Genotype: ACA/ACA<sup>8/11</sup>**FIG. 4B**



10/11

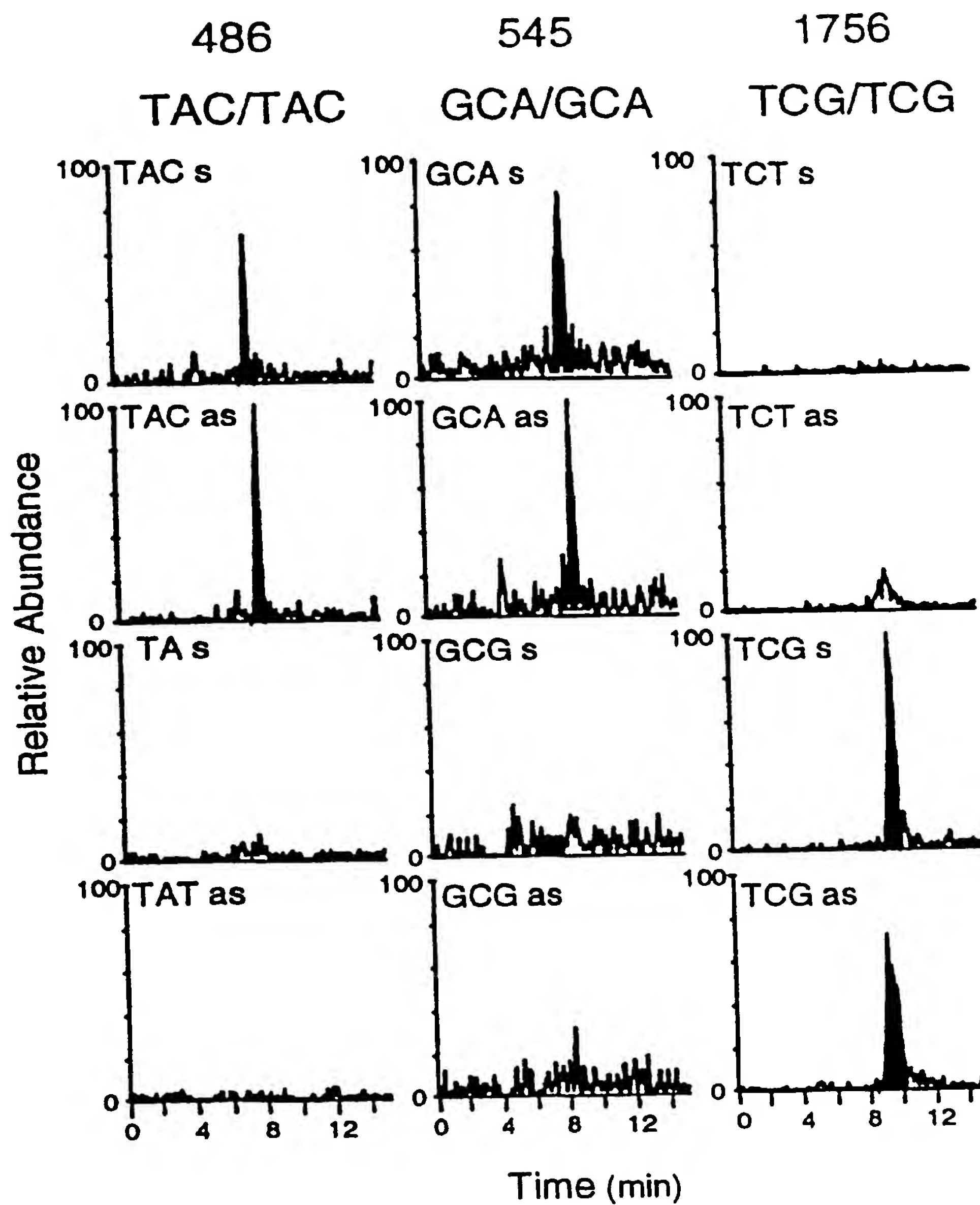
**FIG. 5A**

Codon:        486                    545                    1756  
Genotype: TAT/TAT        GCG/GCG        TCT/TCT



SUBSTITUTE SHEET (RULE 26)

II/II

**FIG. 5B**

SUBSTITUTE SHEET (RULE 26)





## INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

<b>(51) International Patent Classification <sup>7</sup> :</b> <b>C12Q 1/68, G01N 30/72, 33/50</b>	<b>A3</b>	<b>(11) International Publication Number:</b> <b>WO 00/31300</b> <b>(43) International Publication Date:</b> 2 June 2000 (02.06.00)
<b>(21) International Application Number:</b> PCT/US99/27523 <b>(22) International Filing Date:</b> 22 November 1999 (22.11.99) <b>(30) Priority Data:</b> 09/198,340 24 November 1998 (24.11.98) US <b>(71) Applicant (for all designated States except US):</b> THE JOHNS HOPKINS UNIVERSITY [US/US]; Suite 906, 111 Market Place, Baltimore, MD 21202 (US). <b>(72) Inventors; and</b> <b>(75) Inventors/Applicants (for US only):</b> LAKEN, Steven, J. [US/US]; 38 Lawrence Street, Pepperell, MA 01463 (US). VOGELSTEIN, Bert [US/US]; 3700 Breton Way, Baltimore, MD 21208 (US). KINZLER, Kenneth, W. [US/US]; 1403 Halkirk Way, BelAir, MD 21015 (US). GROOPMAN, John, D. [US/US]; 3744 Birchmere Court, Owings Mills, MD 21117 (US). JACKSON, Peta, E. [GB/US]; 721 S. Ann Street, Baltimore, MD 21231 (US). FRIESEN, Marlin, D. [US/US]; 22, rue Benoit Berlioz, Chassieu (FR). <b>(74) Agents:</b> KAGAN, Sarah, A. et al.; Banner & Witcoff, Ltd., 11th floor, 1001 G Street, N.W., Washington, DC 20001-4597 (US).		<b>(81) Designated States:</b> AE, AL, AM, AT, AU, AZ, BA, BB, BG, BR, BY, CA, CH, CN, CR, CU, CZ, DE, DK, DM, EE, ES, FI, GB, GD, GE, GH, GM, HR, HU, ID, IL, IN, IS, JP, KE, KG, KP, KR, KZ, LC, LK, LR, LS, LT, LU, LV, MA, MD, MG, MK, MN, MW, MX, NO, NZ, PL, PT, RO, RU, SD, SE, SG, SI, SK, SL, TJ, TM, TR, TT, TZ, UA, UG, US, UZ, VN, YU, ZA, ZW, ARIPO patent (GH, GM, KE, LS, MW, SD, SL, SZ, TZ, UG, ZW), Eurasian patent (AM, AZ, BY, KG, KZ, MD, RU, TJ, TM), European patent (AT, BE, CH, CY, DE, DK, ES, FI, FR, GB, GR, IE, IT, LU, MC, NL, PT, SE), OAPI patent (BF, BJ, CF, CG, CI, CM, GA, GN, GW, ML, MR, NE, SN, TD, TG).  <b>Published</b> <i>With international search report.</i> <b>(88) Date of publication of the international search report:</b> 12 October 2000 (12.10.00)
<b>(54) Title:</b> GENOTYPING BY MASS SPECTROMETRIC ANALYSIS OF SHORT DNA FRAGMENTS  <b>(57) Abstract</b>  Genotyping can be accomplished by analysis of short, defined DNA segments using electrospray ionization mass spectrometry. The DNA segments are produced using specially designed primers to amplify a cDNA or genomic DNA template. The primers contain a recognition site for a restriction endonuclease. The amplification products are digested with the restriction endonuclease. Single nucleotide polymorphisms can be detected rapidly and reliably.		

**FOR THE PURPOSES OF INFORMATION ONLY**

Codes used to identify States party to the PCT on the front pages of pamphlets publishing international applications under the PCT.

AL	Albania	ES	Spain	LS	Lesotho	SI	Slovenia
AM	Armenia	FI	Finland	LT	Lithuania	SK	Slovakia
AT	Austria	FR	France	LU	Luxembourg	SN	Senegal
AU	Australia	GA	Gabon	LV	Latvia	SZ	Swaziland
AZ	Azerbaijan	GB	United Kingdom	MC	Monaco	TD	Chad
BA	Bosnia and Herzegovina	GE	Georgia	MD	Republic of Moldova	TG	Togo
BB	Barbados	GH	Ghana	MG	Madagascar	TJ	Tajikistan
BE	Belgium	GN	Guinea	MK	The former Yugoslav Republic of Macedonia	TM	Turkmenistan
BF	Burkina Faso	GR	Greece	ML	Mali	TR	Turkey
BG	Bulgaria	HU	Hungary	MN	Mongolia	TT	Trinidad and Tobago
BJ	Benin	IE	Ireland	MR	Mauritania	UA	Ukraine
BR	Brazil	IL	Israel	MW	Malawi	UG	Uganda
BY	Belarus	IS	Iceland	MX	Mexico	US	United States of America
CA	Canada	IT	Italy	NE	Niger	UZ	Uzbekistan
CF	Central African Republic	JP	Japan	NL	Netherlands	VN	Viet Nam
CG	Congo	KE	Kenya	NO	Norway	YU	Yugoslavia
CH	Switzerland	KG	Kyrgyzstan	NZ	New Zealand	ZW	Zimbabwe
CI	Côte d'Ivoire	KP	Democratic People's Republic of Korea	PL	Poland		
CM	Cameroon	KR	Republic of Korea	PT	Portugal		
CN	China	KZ	Kazakhstan	RO	Romania		
CU	Cuba	LC	Saint Lucia	RU	Russian Federation		
CZ	Czech Republic	LI	Liechtenstein	SD	Sudan		
DE	Germany	LK	Sri Lanka	SE	Sweden		
DK	Denmark	LR	Liberia	SG	Singapore		
EE	Estonia						

## INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 99/27523

## A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12Q1/68 G01N30/72 G01N33/50

According to International Patent Classification (IPC) or to both national classification and IPC

## B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12Q G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

## C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	WO 98 12355 A (GENETRACE SYSTEMS ;SHALER THOMAS A (US); TAN YUPING (US); BECKER C) 26 March 1998 (1998-03-26) see whole doc. esp. claims and figures	1-30
X	WO 96 37630 A (STANFORD RES INST INT) 28 November 1996 (1996-11-28) see whole doc. esp. figures and claims, p.20 1.19 ff.	1-30

-/--

☒ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex.

## \* Special categories of cited documents :

- \*A\* document defining the general state of the art which is not considered to be of particular relevance
- \*E\* earlier document but published on or after the international filing date
- \*L\* document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)
- \*O\* document referring to an oral disclosure, use, exhibition or other means
- \*P\* document published prior to the international filing date but later than the priority date claimed

- \*T\* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- \*X\* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- \*Y\* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- \*A\* document member of the same patent family

Date of the actual completion of the international search

19 May 2000

Date of mailing of the international search report

31/05/2000

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2  
NL - 2280 HV Rijswijk  
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,  
Fax: (+31-70) 340-3016

Authorized officer

Müller, F

# INTERNATIONAL SEARCH REPORT

International Application No  
PCT/US 99/27523

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>NORDHOFF E ET AL: "DIRECT MASS SPECTROMETRIC SEQUENCING OF LOW-PICOMOLE AMOUNTS OF OLIGODEOXYNUCLEOTIDES WITH UP TO 21 BASES BY MATRIX-ASSISTED LASER DESORPTION/IONIZATION MASS SPECTROMETRY" JOURNAL OF MASS SPECTROMETRY, GB, WILEY, CHICHESTER, vol. 30, no. 1, 1 January 1995 (1995-01-01), pages 99-112, XP002047878 ISSN: 1076-5174 the whole document</p>	
P, X	<p>LAKEN S.J. ET AL.,: "genotyping by mass spectrometric analysis of short DNA fragments" NATURE BIOTECHNOLOGY, vol. 16, - December 1998 (1998-12) pages 1352-1356, XP002138136 the whole document</p>	1-30

# INTERNATIONAL SEARCH REPORT

information on patent family members

Inter. Application No  
PCT/US 99/27523

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
W0 9812355 A	26-03-1998	US 5965363 A	12-10-1999
		AU 4591697 A	14-04-1998
W0 9637630 A	28-11-1996	US 5700642 A	23-12-1997
		US 5830655 A	03-11-1998
		AU 695705 B	20-08-1998
		AU 5635296 A	11-12-1996
		CA 2220418 A	28-11-1996
		CN 1191575 A	26-08-1998
		DE 69605669 D	20-01-2000
		EP 0828855 A	18-03-1998
		JP 11505127 T	18-05-1999